

cells or MDA-MB-231-Control cells (Figure 1).

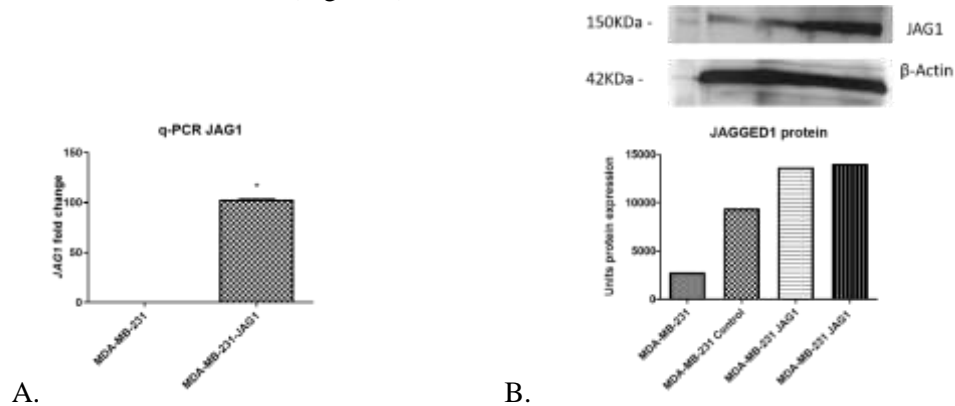


Figure 1: The ligand JAG1 was overexpressed through DNA lentiviral transfection in the breast cancer cells MDA-MB-231. After Blasticin selection, (A) total RNA was extracted and *JAG-1* mRNA levels were evaluated by Real-Time PCR. Relative quantification was done using the $\Delta\Delta C_t$ method normalizing to GAPDH gene expression. Additionally, (B) total protein was isolated and the protein levels of JAG-1 were assessed by Western blot. β -actin was used as a loading control. Data are the mean (S.D.), $n = 3$, $*p < 0.05$, by two-tailed unpaired Student's t-test.

MDA-MB-231-JAG1 or MDA-MB-231 cells were next subcutaneously injected in Balb/c nude mice. We could observe that the overexpression of JAG1 significantly increases the growth of MDA-MB-231-JAG1 cells when compared to MDA-MB-231-Control cells (Figure 2).

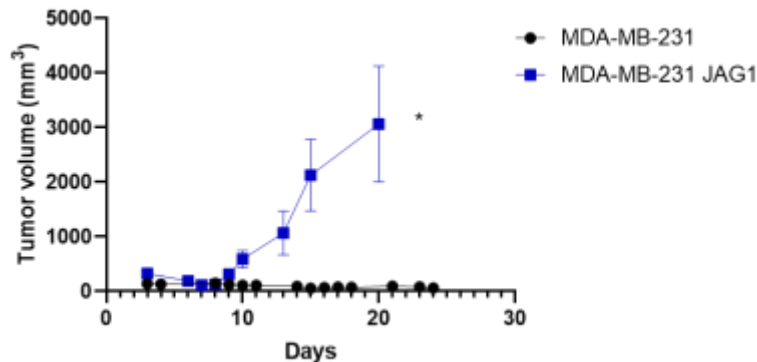


Figure 2: Tumor growth curve showing the tumor volume of MDA-MB-231-Control and MDA-MB-231-JAG1 cells. Data are the mean (S.D.), $n = 5$. Data were analyzed by non-linear regression. $*p < 0.05$, by two-tailed unpaired Student's t-test.

The SELEX assay was done using MDA-MB-231-Control cells as a negative control cycle and MDA-MB-231-JAG1 as a positive control cycle. All positive cycles had a good extraction efficiency with a yield of 248.9 ng/ul in the third cycle (Table 1). The negative cycles were discarded and positives cycles were stored for further purification and characterization.

Table 1: SELEX aptamer cycle.

Cycle – SELEX	N° of cells	tRNA (25ug/ul)	Washing with selection buffer 1x	Pool Concentration	Incubation Time	Quantification
1	10 ⁶	0	0	100 uM	40min	288,5ng/ul
2	10 ⁵	1ul	3x	10 uM	40min	46,1ng/ul
3	5 x 10 ⁴	2ul	6x	10 uM	30min	248,9ng/ul

The immunofluorescence assay showed that the selected aptamers against MDA-MB-231JAG1 cells from the third cycle of SELEX (Table 1) were able to bind MDA-MB-231-JAG1 cells more specifically than the selected aptamer from the first cycle of SELEX or the aptamer library. Besides that, at 40-fold magnification, we also found that aptamer binds to cell membrane, cell cytoplasm and small vesicles within cells (Figure 3).

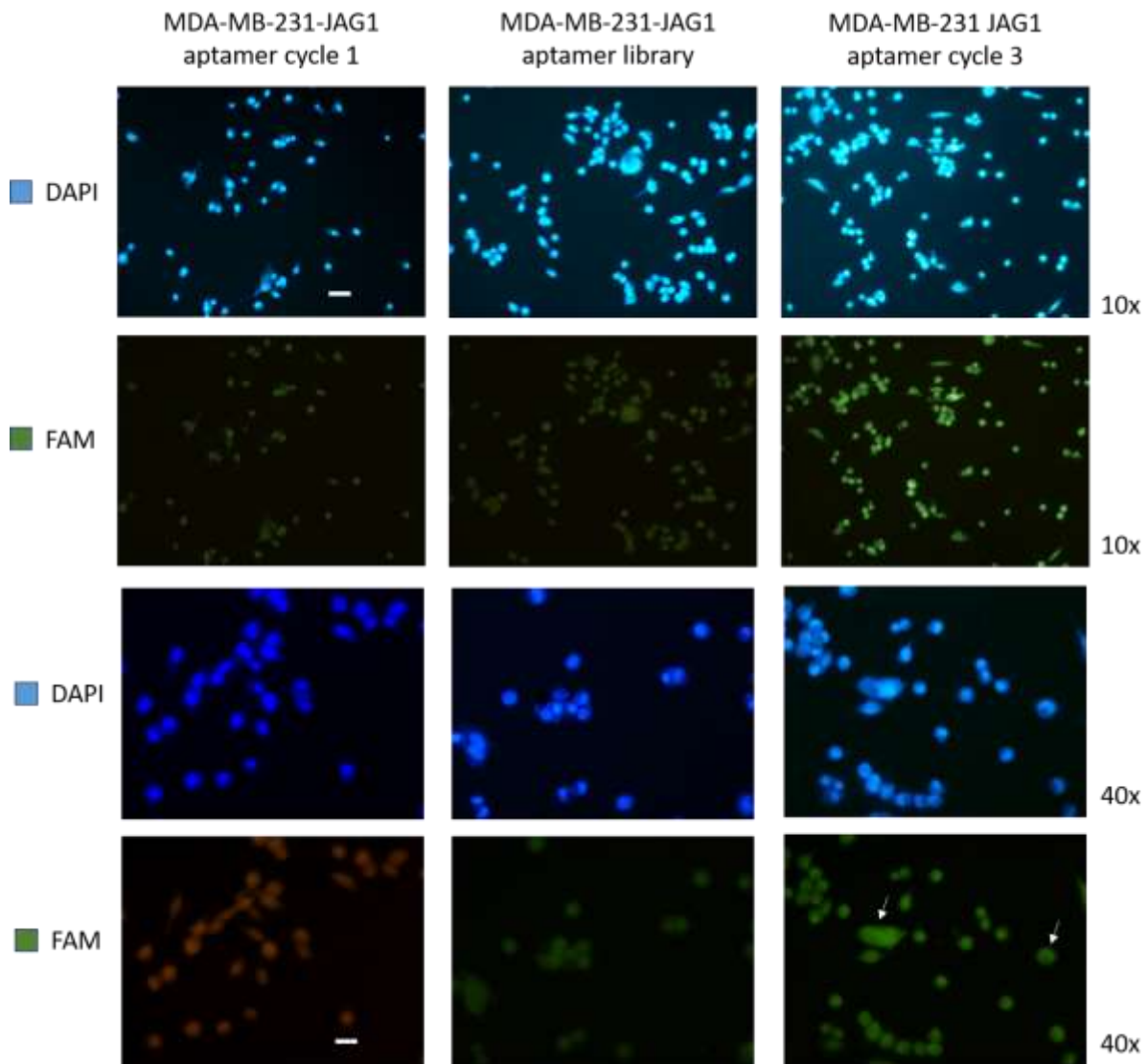


Figure 3: MDA-MB-231-JAG1 in a monolayer were fixed in 4% PFA and then FAM-aptamers from cycle 1, 3 or the library were incubated with cells. Representative immunofluorescence images (10X or 40X magnification) showing JAG1 aptamers (green) and nucleus (blue) staining are presented. White arrow shows cytoplasmic vesicles. Scale bar, 200 μ m for 10x and 800 μ m for 40x of magnification.

Altogether our results demonstrate the selection of a new molecule based on ssDNA aptamers able to specifically bind to breast cancer cells that overexpress JAG1 protein. This aptamer has the potential to be radiolabeled with diagnostic or therapeutic radionuclides (eg ^{99m}Tc , ^{18}F or ^{177}Lu). Indeed, several studies have demonstrated that aptamers have the potential to be used as therapeutic or diagnostic agents as AS1411 and MAG3-apt radiopharmaceuticals through their radiolabeling with radionuclides and their affinity with molecular targets [6]–[8].

4. Conclusions

Our work showed that MDA-MB-231-JAG1 cells overexpress more mRNA and JAG1 protein than control cells (MDA-MB-231-Control), being an important tool for the study of JAG1 biology in tumors. We also selected aptamers with high affinity for MDA-MB-231-JAG1 cells that could be a useful tool for the development of new radiopharmaceutical for the diagnosis and treatment of tumors that overexpress JAG1.

Acknowledgements

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